

# Temporal Exponential Random Graph Models (TERGMs) for dynamic network modeling in statnet

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[http://statnet.csde.washington.edu/statnet\\_users\\_group.shtml](http://statnet.csde.washington.edu/statnet_users_group.shtml)

## 1. Getting Started

Open an R session, and set your working directory to the location where you would like to save this work.

To install all of the packages in the statnet suite:

```
install.packages('statnet')  
library(statnet)
```

Or, to only install the specific statnet packages needed for this tutorial:

```
install.packages('tergm') # will install the network package
install.packages('sna')
```

After the first time, to update the packages one can either repeat the commands above, or use:

```
update.packages('name.of.package')
```

For this tutorial, we will need one additional package (coda), which is recommended (but not required) by ergm:

```
install.packages('coda')
```

Make sure the packages are attached:

```
library(statnet)
```

or

```
library(tergm)
library(sna)
library(coda)
```

Check package version

```
# latest versions:  tergm 3.4.0, ergm 3.6.0, network 1.13.0, networkDynamic 0.9.0 (as of 4/2/2016)
sessionInfo()
```

Set seed for simulations – this is not necessary, but it ensures that we all get the same results (if we execute the same commands in the same order).

```
set.seed(0)
```

## 2. A quick review of static ERGMs

Exponential-family random graph models (ERGMs) represent a general class of models based in exponential-family theory for specifying the probability distribution underlying a set of random graphs or networks. Within this framework, one can—among other tasks—obtain maximum-likelihood estimates for the parameters of a specified model for a given data set; simulate additional networks with the underlying probability distribution implied by that model; test individual models for goodness-of-fit, and perform various types of model comparison.

The basic expression for the ERGM class can be written as:

$$P(Y = y) = \frac{\exp(\theta'g(y))}{k(\theta)}$$

where:

- $Y$  is the random variable for the state of the network (with realization  $y$ )
- $g(y)$  is the vector of model statistics for network  $y$ ,

- $\theta$  is the vector of coefficients for those statistics
- $k(\theta)$  represents the quantity in the numerator summed over all possible networks (typically constrained to be all networks with the same node set as  $y$ ).

This can be re-expressed in terms of the conditional log-odds of a single actor pair:

$$\text{logit } P(Y_{ij} = 1 \mid y_{ij}^c) = \ln \frac{P(Y_{ij}=1|y_{ij}^c)}{P(Y_{ij}=0|y_{ij}^c)} = \theta' \delta(g(y))_{ij}$$

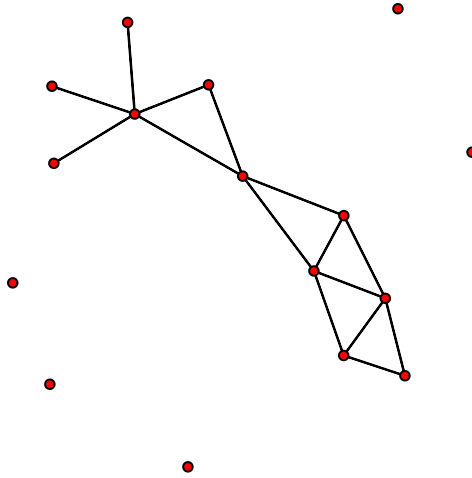
where:

- $Y_{ij}$  is the random variable for the state of the actor pair  $i, j$  (with realization  $y_{ij}$ )
- $y_{ij}^c$  signifies the complement of  $y_{ij}$ , i.e. all dyads in the network other than  $y_{ij}$ .
- $\delta(g(y))_{ij}$  equals  $g(y_{ij}^+) - g(y_{ij}^-)$ , where
- $y_{ij}^+$  is defined as  $y_{ij}^c$  along with  $y_{ij}$  set to 1
- $y_{ij}^-$  is defined as  $y_{ij}^c$  along with  $y_{ij}$  set to 0.
- That is,  $\delta(g(y))_{ij}$  equals the value of  $g(y)$  when  $y_{ij} = 1$  minus the value of  $g(y)$  when  $y_{ij} = 0$ , but all other dyads are as in  $g(y)$ . This emphasizes the log-odds of an individual tie conditional on all others.
- We call  $g(y)$  the *statistics* of the model, and  $\delta(g(y))_{ij}$  the *change statistics* for actor pair  $y_{ij}$ .

Fitting an ERGM usually begins with obtaining data:

```
data("florentine")
ls()
```

```
plot(flobusiness)
```



To refresh our memories on ERGM syntax, let us fit a cross-sectional example. Just by looking at the plot of `flobusiness`, we might guess that there are more triangles than expected by chance for a network of this size and density, and thus that there is some sort of explicit triangle closure effect going on. One useful way to model this effect in ERGMs that has been explored in the literature is with a `gwesp` statistic. When the alpha parameter of this statistic equals 0, then the statistic counts the number of edges in the network that are in at least one triangle. You can see this by summarizing this network and then counting them for yourself in the visualization:

```
summary(flobusiness~edges+gwesp(0, fixed=T))
```

```
edges gwesp.fixed.0
    15          12
```

```
fit1 <- ergm(flobusiness~edges+gwesp(0, fixed=T))
```

Starting maximum likelihood estimation via MCMLE:

Iteration 1 of at most 20:

The log-likelihood improved by 0.3362

Step length converged once. Increasing MCMC sample size.

Iteration 2 of at most 20:

The log-likelihood improved by 0.01704

Step length converged twice. Stopping.

Evaluating log-likelihood at the estimate. Using 20 bridges: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17

This model was fit using MCMC. To examine model diagnostics and check for degeneracy, use the `mcmc.diag`

```
summary(fit1)
```

```
=====
Summary of model fit
=====

Formula:   flobusiness ~ edges + gwesp(0, fixed = T)

Iterations: 2 out of 20

Monte Carlo MLE Results:
      Estimate Std. Error MCMC % p-value
edges      -3.3626    0.6267    0 < 1e-04 ***
gwesp.fixed.0  1.5668    0.5927    0 0.00933 **
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

      Null Deviance: 166.36 on 120 degrees of freedom
      Residual Deviance: 78.34 on 118 degrees of freedom

AIC: 82.34   BIC: 87.92   (Smaller is better.)
```

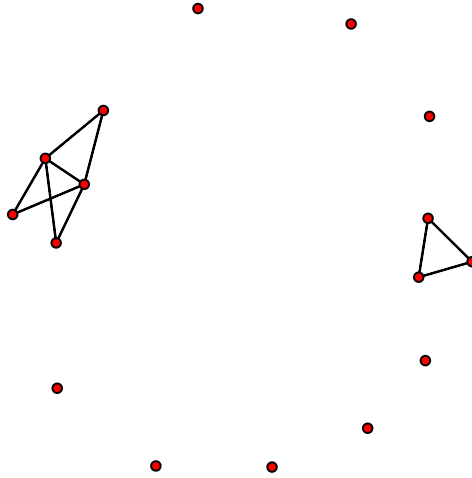
How to interpret the coefficients? Conditional on the rest of the network, a given edge's log-odds of existing depends on how much its presence changes the number of edges in at least one triangle in the network.

- If the answer is 0 (i.e. the two incident nodes have no relational partners in common), the log-odds are  $-3.36$ .
- If the answer is 1 then the log-odds are  $-3.36 + 1.57 = -1.79$ .
- If the answer is 2 then the log-odds are  $-3.36 + 1.57 \times 2 = -0.220$ .
- The corresponding probabilities are 3.4%, 14.3%, and 44.5%.

How can adding one edge change the number of edges in at least one triangle by 2?

With the estimation in place, we can simulate a new network from the given model:

```
sim1 <- simulate(fit1,nsim=1,
                 control=control.simulate.ergm(MCMC.burnin=1e5))
plot(sim1)
```



### 3. An Introduction to STERGMs (non-technical)

- Separable Temporal ERGMs (STERGMs) are an extension of ERGMs for modeling dynamic networks in discrete time, introduced in Krivitsky and Handcock (2010).
- The cross-sectional ERGM entails a single network, and a single model on that network. STERGMs, in contrast, posit two models: one ERGM underlying relational formation, and a second one underlying relational dissolution. Specifying a STERGM thus entails writing two ERGM formulas instead of one.
- STERGMs also requires dynamic data, of course; such data can come in many forms, and we will cover a few examples today.
- This approach is not simply a methodological development, but a theoretical one as well, and one which matches common sense for many social processes. Think of romantic relations. It seems intuitive that the statistical model underlying relational formation (i.e. affecting who becomes partners with whom, out of the set of people who aren't already) is likely to be different than the model underlying relational dissolution (i.e. affecting who breaks up with whom, out of the set of people currently in relationships). Any reasonable model of the former would need to include a variety of homophily parameters (mixing on age, for example). The latter may or may not. (Conditional on being in a relationship, does your difference in age affect your probability of breaking up? Perhaps, but probably not as fundamentally or as strongly as for formation).
- In thinking about how STERGMs work, and how they relate to different data forms, it can be helpful to think of the approximation commonly used in epidemiology to consider basic disease dynamics:

$$\text{prevalence} = \text{incidence} \times \text{duration}$$

- To translate this to dynamic networks: first think of a Bernoulli model where all ties are equally likely. This expression indicates that the number of ties present in the cross-section (prevalence) is a function of the rate at which they form (incidence) and the rate at which they dissolve (1/duration). The faster ties form, the more ties that are present in the cross-section. And the slower ties dissolve, the more ties that are present in the cross-section.
- This same concept extends to more elaborate models beyond just Bernoulli. Imagine a model with triangles in it—the faster that triangles are formed and the slower that they’re dissolved, the more that will be present in the cross-section.
- Using the notation from the ERGM review above: the two equations governing a STERGM are commonly called the formation equation and the dissolution (or persistence) equation, respectively:

$$\ln \frac{P(Y_{ij,t+1}=1|y_{ij}^c, Y_{ij,t}=0)}{P(Y_{ij,t+1}=0|y_{ij}^c, Y_{ij,t}=0)} = \theta^+ \delta(g^+(y))_{ij}$$

$$\ln \frac{P(Y_{ij,t+1}=1|y_{ij}^c, Y_{ij,t}=1)}{P(Y_{ij,t+1}=0|y_{ij}^c, Y_{ij,t}=1)} = \theta^- \delta(g^-(y))_{ij}$$

- These largely parallel the one ERGM equation. We have simply:
- added a time index to the tie values
- added in a new conditional. In the formation equation, the expression is conditional on the tie *not* existing at the prior time step, and in the dissolution equation it is conditional on the tie existing; and
- defined two  $\theta$  and  $g$  vectors instead of just one. Now there are  $\theta^+$  and  $g^+(y)_{ij}$ , reflecting the coefficients and statistics in the formation model, and  $\theta^-$  and  $g^-(y)_{ij}$  reflecting those in the dissolution model.
- Notice that for the dissolution model, the  $P(Y_{ij,t+1} = 1)$  is in the numerator and the  $P(Y_{ij,t+1} = 0)$  is in the denominator. This is needed to make the mathematics parallel to that of the formation model; however, it is also what causes the model to be an expression of relational persistence rather than of dissolution. Of course, the probability of dissolution is merely 1 - persistence, so the model captures both phenomena, and conceptually we often think about relational dissolution in parallel with formation.
- To understand the implications of the two  $\theta$  and  $g$  vectors, imagine a model for romantic relationships that captures propensities for:
  - people who are currently in one relationship to not form a new relationship as often as single people do; and
  - those few people who are in two ongoing relationships to dissolve one of them more often than others do.
- Both the formation and dissolution (persistence) equations would contain the ergm term “concurrent”, which counts the number of nodes in the network of degree 2 or more.
- In both models, the coefficient for the concurrent term would be negative; new ties are relatively unlikely to form if they will generate a new node of degree 2 or more. And they are relatively unlikely to persist if they maintain the current number of nodes of degree 2 or more, when they could bring it down by dissolving.
- Finally, remember that the  $s$  in *stergm* stands for separable; this refers to the fact that the two models assume that relational formation and dissolution occur independently of one another within a time step. We explore how this works and its implications in the examples.

## 4. An Introduction to STERGMs (more formal)

To understand STERGMs formally, we first review the ERGM framework for cross-sectional or static networks again, this time with more complete notation. Let  $\mathbb{Y} \subseteq \{1, \dots, n\}^2$  be the set of potential relations (dyads) among  $n$  nodes, ordered for directed networks and unordered for undirected. We can represent a network  $\mathbf{y}$  as a set of ties, with the set of possible sets of ties,  $\mathcal{Y} \subseteq 2^{\mathbb{Y}}$ , being the sample space:  $\mathbf{y} \in \mathcal{Y}$ . Let  $\mathbf{y}_{ij}$  be 1 if  $(i, j) \in \mathbf{y}$  — a relation of interest exists from  $i$  to  $j$  — and 0 otherwise.

The network also has an associated covariate array  $\mathbf{X}$  containing attributes of the nodes, the dyads, or both. An exponential-family random graph model (ERGM) represents the pmf of  $\mathbf{Y}$  as a function of a  $p$ -vector of network statistics  $g(\mathbf{Y}, \mathbf{X})$ , with parameters  $\theta \in \mathbb{R}^p$ , as follows:

$$\Pr(\mathbf{Y} = \mathbf{y} \mid \mathbf{X}) = \frac{\exp\{\theta \cdot g(\mathbf{y}, \mathbf{X})\}}{k(\theta, \mathbf{X}, \mathcal{Y})},$$

where the normalizing constant  $k(\theta, \mathbf{X}, \mathcal{Y}) = \sum_{\mathbf{y}' \in \mathcal{Y}} \exp\{\theta \cdot g(\mathbf{y}', \mathbf{X})\}$  is a summation over the space of possible networks on  $n$  nodes,  $\mathcal{Y}$ . Where  $\mathcal{Y}$  and  $\mathbf{X}$  are held constant, as in a typical cross-sectional model, they may be suppressed in the notation. Here, on the other hand, the dependence on  $\mathcal{Y}$  and  $\mathbf{X}$  is made explicit.

In modeling the transition from a network  $\mathbf{Y}^t$  at time  $t$  to a network  $\mathbf{Y}^{t+1}$  at time  $t+1$ , the separable temporal ERGM assumes that the formation and dissolution of ties occur independently from each other within each time step, with each half of the process modeled as an ERGM. For two networks (sets of ties)  $\mathbf{y}, \mathbf{y}' \in \mathcal{Y}$ , let  $\mathbf{y} \supseteq \mathbf{y}'$  if any tie present in  $\mathbf{y}'$  is also present in  $\mathbf{y}$ . Define  $\mathcal{Y}^+(\mathbf{y}) = \{\mathbf{y}' \in \mathcal{Y} : \mathbf{y}' \supseteq \mathbf{y}\}$ , the networks that can be constructed by forming ties in  $\mathbf{y}$ ; and  $\mathcal{Y}^-(\mathbf{y}) = \{\mathbf{y}' \in \mathcal{Y} : \mathbf{y}' \subseteq \mathbf{y}\}$ , the networks that can be constructed dissolving ties in  $\mathbf{y}$ .

Given  $\mathbf{y}^t$ , a formation network  $\mathbf{Y}^+$  is generated from an ERGM controlled by a  $p$ -vector of formation parameters  $\theta^+$  and formation statistics  $g^+(\mathbf{y}^+, \mathbf{X})$ , conditional on only adding ties:

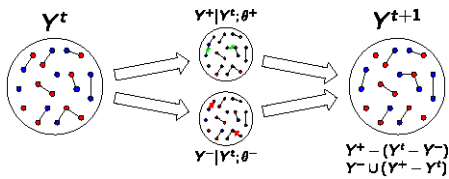
$$\Pr(\mathbf{Y}^+ = \mathbf{y}^+ \mid \mathbf{Y}^t; \theta^+) = \frac{\exp\{\theta^+ \cdot g^+(\mathbf{y}^+, \mathbf{X})\}}{k(\theta^+, \mathbf{X}, \mathcal{Y}^+(\mathbf{Y}^t))}, \quad \mathbf{y}^+ \in \mathcal{Y}^+(\mathbf{y}^t).$$

A dissolution network  $\mathbf{Y}^-$  is simultaneously generated from an ERGM controlled by a (possibly different)  $q$ -vector of dissolution parameters  $\theta^-$  and corresponding statistics  $g^-(\mathbf{y}^-, \mathbf{X})$ , conditional on only dissolving ties from  $\mathbf{y}^t$ , as follows:

$$\Pr(\mathbf{Y}^- = \mathbf{y}^- \mid \mathbf{Y}^t; \theta^-) = \frac{\exp\{\theta^- \cdot g^-(\mathbf{y}^-, \mathbf{X})\}}{k(\theta^-, \mathbf{X}, \mathcal{Y}^-(\mathbf{Y}^t))}, \quad \mathbf{y}^- \in \mathcal{Y}^-(\mathbf{y}^t).$$

The cross-sectional network at time  $t+1$  is then constructed by applying the changes in  $\mathbf{Y}^+$  and  $\mathbf{Y}^-$  to  $\mathbf{y}^t$ , as follows:  $\mathbf{Y}^{t+1} = \mathbf{Y}^t \cup (\mathbf{Y}^+ - \mathbf{Y}^t) - (\mathbf{Y}^t - \mathbf{Y}^-)$ . which simplifies to either:  $\mathbf{Y}^{t+1} = \mathbf{Y}^+ - (\mathbf{Y}^t - \mathbf{Y}^-)$  or  $\mathbf{Y}^{t+1} = \mathbf{Y}^- \cup (\mathbf{Y}^+ - \mathbf{Y}^t)$

Visually, we can sum this up as:



## 5. Notes on model specification and syntax

Within `statnet`, an ERGM involves one network and one set of network statistics, so these are specified together using R's formula notation:

```
my.network ~ my.vector.of.model.terms
```

For a call to `stergm`, there is still one network (or a list of networks), but two formulas. These are now passed as three separate arguments: the network(s) (argument `nw`), the formation formula (argument `formation`),



and the dissolution formula (argument `dissolution`). The latter two both take the form of a one-sided formula. E.g.:

```
stergm(my.network,
       formation= ~edges+kstar(2),
       dissolution= ~edges+triangle
       )
```

There are other features of a call to either `ergm` or `stergm` that will be important; we list these features here, and illustrate them in one or more examples during the workshop.

- To fix the coefficient for a particular network statistic, one uses offset notation. For instance, to fix a dissolution model with only an edges term with parameter value 4.2, the dissolution formula would be:

```
dissolution = ~offset(edges)
```

and the corresponding argument for passing the parameter value would be:

```
offset.coef.diss = 4.2
```

- In parallel with `ergm`, any information used to specify the nature of the fitting algorithm is passed by specifying a vector called `control.stergm` to the `control` argument. For example:

```
control=control.stergm(MCMC.burnin=10000)
```

For a list of options, type `?control.stergm`

- Another argument that the user must supply is `estimate`, which controls the estimation method. Unlike with cross-sectional ERGMs, there is not necessarily an obvious default here, as different scenarios are best fit with different approaches. The most important for the new user to recognize are `EGMME` (equilibrium generalized method of moments estimation) and `CMLE` (conditional maximum likelihood estimation). A good rule of thumb is that when fitting to two (or more) networks (i.e. with panel data), one should use `estimate="CMLE"`; and when fitting to a single cross-section with some duration information, use `estimate="EGMME"`.

## 6. Example 1: Multiple network cross-sections

One common form of data for modeling dynamic network processes consists of observations of network structure at two or more points in time on the same node set. Many classic network studies were of this type, and data of this form continue to be collected and analyzed.

Let us consider the famous Sampson monastery data:

```
data(samplk)
ls()
```

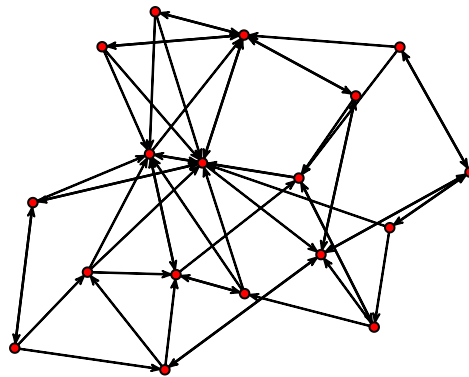
```
[1] "fit1"          "flobusiness" "flomarriage" "samplk1"      "samplk2"
[6] "samplk3"      "sim1"
```

To pass them into `stergm`, we need to combine them into a list:

```
samp <- list()
samp[[1]] <- samplk1
samp[[2]] <- samplk2
samp[[3]] <- samplk3
```

Now we must decide on a model to fit to them. Plotting one network:

```
plot(samplk1)
```



we might get the idea to consider mutuality as a predictor of a directed edge. Also, since this is a directed network, and there appear to be a considerable number of triadic relations, it might be worth investigating the role of cyclicity vs. transitivity in the network. Although there are a number of ways to model this latter question, we will select the relatively robust measures `transitiveties` and `cyclicalties` (the number of ties  $i \rightarrow j$  that have at least one two-path from  $i \rightarrow j$  and from  $j \rightarrow i$ , respectively).

Since we have multiple network snapshots, and we have separate formation and dissolution models, we can estimate the degree to which closing a mutual dyad or closing a triad of each type predicts the creation of a tie, and also estimate the degree to which maintaining a mutual dyad or maintaining a triad of each type predicts the persistence of an existing tie. We might see different phenomena at work in each case; or the same phenomena, but with different coefficients.

In the following code, we pass five arguments: the series of networks that we want to model (`samp`), the formation formula, the dissolution formula, the fitting algorithm (in this case, conditional MLE is best since we have a network time series), and the list of time slices we wish to model (which includes all in the passed network series by default):

```
samp.fit <- stergm(samp,
  formation= ~edges+mutual+cyclicalities+transitiveties,
  dissolution = ~edges+mutual+cyclicalities+transitiveties,
  estimate = "CMLE",
  times=1:3
)
```

Fitting formation:

Iteration 1 of at most 20:

=====  
Lots of output snipped  
=====

This model was fit using MCMC. To examine model diagnostics and check for degeneracy, use the `mcmc.diag`

And the results:

```
summary(samp.fit)
```

```
=====
Summary of formation model fit
=====
```

Formula: ~edges + mutual + cyclicalities + transitiveties

Iterations: 4 out of 20

Monte Carlo MLE Results:

	Estimate	Std. Error	MCMC %	p-value
edges	-3.4663	0.3384	0	<1e-04 ***
mutual	2.0431	0.4028	0	<1e-04 ***
cyclicalities	-0.1397	0.2025	0	0.4906
transitiveties	0.3942	0.2388	0	0.0995 .

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Null Deviance: 693.1 on 500 degrees of freedom

Residual Deviance: 240.2 on 496 degrees of freedom

AIC: 248.2 BIC: 265.1 (Smaller is better.)

```
=====
Summary of dissolution model fit
=====
```

Formula: ~edges + mutual + cyclicalities + transitiveties

Iterations: 2 out of 20

Monte Carlo MLE Results:

	Estimate	Std. Error	MCMC %	p-value
edges	0.2013	0.2975	0	0.5002

```

mutual      0.7926    0.5134    0  0.1255
cyclicalties -0.1870    0.2522    0  0.4600
transitiveties 0.5057    0.2812    0  0.0749 .
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Null Deviance: 155.3 on 112 degrees of freedom
Residual Deviance: 136.7 on 108 degrees of freedom

AIC: 144.7    BIC: 155.6    (Smaller is better.)

```

So, all else being equal, a relationship is much more likely to form if it will close a mutual pair; the log-odds are increased by 2.04. Note that the most that the change statistic can equal in this case is 1.

The effect on persistence is also positive, but less strong and clear; the point estimate is an increase of 0.79 in the log-odds of persistence.

Transitivity may also matter, perhaps slightly more so in terms of dissolution than formation.

If one wishes to include only a subset of times from one's network series, one can alter the `times` argument:

```

samp.fit.2 <- stergm(samp,
  formation= ~edges+mutual+cyclicalties+transitiveties,
  dissolution = ~edges+mutual+cyclicalties+transitiveties,
  estimate = "CMLE",
  times=1:2
)

```

How do these coefficients compare? How about the standard errors?

## 7. Example 2: One network cross section and durational information

Now, let us imagine a different scenario in which we have observed two types of data: a single cross-sectional network, and a mean relational duration. Let us say the cross-sectional network is `flobusiness`, and the mean relational duration we have witnessed is 10 time steps. Furthermore, we are willing (for reasons of theory or convenience) to assume a purely homogeneous dissolution process (that is, every existing relationship has the same probability of dissolving as all others, and at all times).

For a cross-sectional ERGM, a purely homogeneous model is one with just a single term in it for an edge count. The same is true for either of the two formulas in a STERGM.

**First**, we specify formation and dissolution models (`formation` and `dissolution`). We will begin by assuming a formation model identical to the model we fit in the cross-sectional case:

```
formation = ~edges+gwesp(0, fixed=T)
```

Analogously to cross-sectional ERGMs, our assumption of completely homogeneous dissolution corresponds to a model with only an edgcount term in it. As we will see in the next step, however, we are going to supply the `stergm` algorithm with the coefficient for our dissolution model, and not have the algorithm try to estimate it. In STERGM notation this is:

```
dissolution = ~offset(edges)
```

**Second**, we calculate `theta.diss`. Our dissolution model is applied to the set of ties that exist at any given time point, as reflected in the conditional present in both the numerator and denominator:

$$\ln \frac{P(Y_{ij,t+1}=1|Y_{ij,t}=1)}{P(Y_{ij,t+1}=0|Y_{ij,t}=1)} = \theta' \delta(g(y))_{ij}$$

Recall that the numerator represents the case where a tie persists to the next step, and the denominator represents the case where it dissolves.

Furthermore, the one term in our  $\delta(g(y))_{ij}$  vector is the change statistic on network edge count, which equals one for all  $i, j$ .

We define the probability of persistence from one time step to the next for actor pair  $i, j$  as  $p_{ij}$ , and the probability of dissolution as  $q_{ij} = 1 - p_{ij}$ . Our dissolution model is Bernoulli; that is, all edges have the same probability of dissolution, and thus of persistence, so we further define  $p_{ij} = p$  for all  $i, j$  and  $q_{ij} = q$  for all  $i, j$ . Then:

$$\ln\left(\frac{p_{ij}}{1-p_{ij}}\right) = \theta' \delta(g(y))_{ij}$$

$$\ln\left(\frac{p}{1-p}\right) = \theta$$

$$\ln\left(\frac{1-q}{q}\right) = \theta$$

$$\ln\left(\frac{1}{q} - 1\right) = \theta$$

And because this is a discrete memoryless process, durations are geometric; and for the geometric distribution, the mean is equal to the reciprocal of the dissolution probability. Symbolizing mean relational duration as  $d$ , we have  $d = \frac{1}{q}$ , and thus:

$$\theta = \ln(d - 1)$$

So, for our dissolution model,  $\text{theta.diss} = \ln(10 - 1) = \ln 9 = 2.197$ :

```
theta.diss <- log(9)
```

In short, because our dissolution model is dyadic independent, we can calculate it using a (rather simple) closed form solution.

**Third**, we decide upon our targets. For cross-sectional ERGMs, the model is by default fit using the sufficient statistics present in the cross-sectional network. For the STERGM with two cross-sections that we fit above, we also wanted to use the sufficient statistics present in the two observed networks. In the case of one cross-section but a formation and dissolution model, the reasonable default is less clear. Thus, the user is required to supply this information via the `targets` argument. This can take a one-sided formula listing the terms to be fit using the data in the network cross-section; or, if the formula is identical to either the formation or dissolution model, the user can simply pass the string `"formation"` or `"dissolution"`, respectively. If one is specifying `targets="formation"`, dissolution should be an offset, and vice versa. If the values to be targeted for those terms are anything other than the sufficient statistics present in the cross-sectional network, then those values can be passed with the argument `target.stats`. In this case, we want to use the sufficient statistics present in the cross-sectional network for the model terms in the formation model.

**Finally**, we estimate the formation model, conditional on the dissolution model. We put it all together for our first call to `stergm`, adding in one additional control argument that helps immensely with monitoring model convergence (and is just plain cool): plotting the progress of the coefficient estimates and the simulated sufficient statistics in real time. When one is using a GUI tool like RStudio, it helps to output this plotting to a separate window, which we do below with the function `X11` (and then turn off that window again with `dev.off()`):

```
X11()
stergm.fit.1 <- stergm(flobusiness,
  formation= ~edges+gwesp(0, fixed=T),
  dissolution = ~offset(edges),
  targets="formation",
  offset.coef.diss = theta.diss,
  estimate = "EGMME",
```

```
control=control.stergm(SA.plot.progress=TRUE)
)
dev.off()
```

Iteration 1 of at most 20:

=====  
===== Lots of output snipped =====

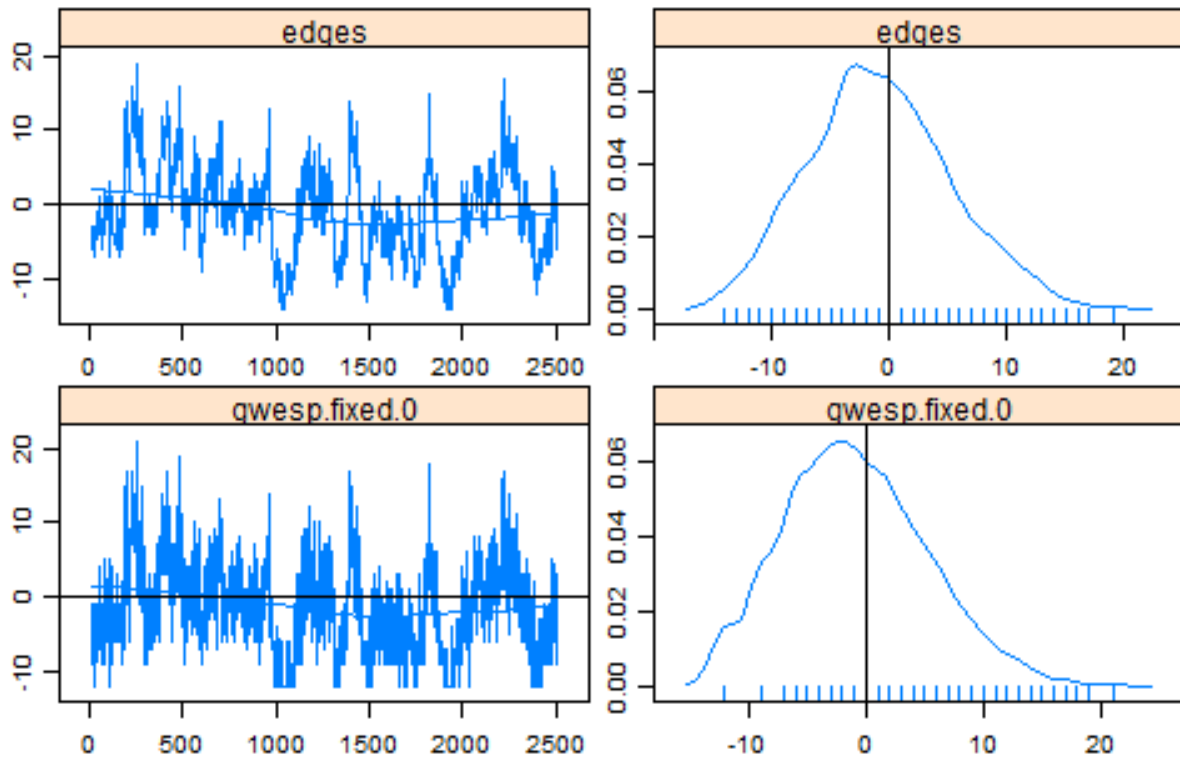
== Phase 3: Simulate from the fit and estimate standard errors.==

The real-time feedback suggests that the fitting went well, but let's double-check:

```
mcmc.diagnostics(stergm.fit.1)
```

```
=====
EGMME diagnostics
=====
=====  
===== Lots of output snipped =====
```

## Sample statistics



Since these look good, we can next query the object in a variety of ways to see what we have:

```
stergm.fit.1
```

```
Formation Coefficients:
  edges qwesp.fixed.0
-6.649    2.447
Dissolution Coefficients:
edges
2.197
```

```
names(stergm.fit.1)
```

```
[1] "network"      "formation"    "dissolution"
```

```
[4] "targets"      "target.stats"  "estimate"
[7] "covar"        "opt.history"   "sample"
[10] "sample.obs"   "control"       "reference"
[13] "mc.se"        "constraints"   "formation.fit"
[16] "dissolution.fit"
```

```
stergm.fit.1$formation
```

```
~edges + gwesp(0, fixed = T)
```

```
stergm.fit.1$formation.fit
```

```
EGMME Coefficients:
```

```
      edges gwesp.fixed.0
-6.649      2.447
```

```
summary(stergm.fit.1)
```

```
=====
Summary of formation model fit
=====
```

```
Formula:    ~edges + gwesp(0, fixed = T)
```

```
Iterations: NA
```

```
Equilibrium Generalized Method of Moments Results:
```

	Estimate	Std. Error	MCMC %	p-value
edges	-6.6494	0.6985	0	<1e-04 ***
gwesp.fixed.0	2.4471	0.9384	0	0.0103 *

```
---
```

```
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
=====
Summary of dissolution model fit
=====
```

```
Formula:    ~offset(edges)
```

```
Iterations: NA
```

```
Equilibrium Generalized Method of Moments Results:
```

	Estimate	Std. Error	MCMC %	p-value
edges	2.197	0.000	0	<1e-04 ***

```
---
```

```
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
The following terms are fixed by offset and are not estimated:
```

```
edges
```



We have now obtained estimates for the coefficients of a formation model that, conditional on the stated dissolution model, yields simulated targets that matched those observed. Something very useful we have also gained in the process is the ability to simulate networks with the desired cross-sectional structure and mean relational duration. This ability serves us well for any application areas that requires us to simulate phenomena on dynamic networks, whether they entail the diffusion of information or disease, or some other process.

```
stergm.sim.1 <- simulate.stergm(stergm.fit.1, nsim=1,  
  time.slices = 1000)
```

Understanding this object requires us to learn about an additional piece of `statnet` functionality: the `networkDynamic` package.

## 8. networkDynamic

In `statnet`, cross-sectional networks are stored using objects of class `network`. Tools to create, edit, and query network objects are in the package `network`. Dynamic networks are now stored as objects with two classes (`network` and `networkDynamic`). They can thus be edited or queried using standard functions from the `network` package, or using additional functions tailored specifically to the case of dynamic networks in the package `networkDynamic`.

To illustrate, let us begin with the network that we just created:

```
stergm.sim.1
```

NetworkDynamic properties:

```
  distinct change times: 921  
  maximal time range: -Inf until  Inf
```

Includes optional `net.obs.period` attribute:

```
Network observation period info:  
  Number of observation spells: 2  
  Maximal time range observed: 0 until 1001  
  Temporal mode: discrete  
  Time unit: step  
  Suggested time increment: 1
```

Network attributes:

```
vertices = 16  
directed = FALSE  
hyper = FALSE  
loops = FALSE  
multiple = FALSE  
bipartite = FALSE  
net.obs.period: (not shown)  
total edges= 120  
  missing edges= 0  
  non-missing edges= 120
```

Vertex attribute names:

```
priorates totalties vertex.names wealth
```

Edge attribute names:

```
active
```

We can deduce from the number of edges that this likely represents the cumulative network—that is, the union of all edges that exist at any point in time over the course of the simulation. This is because the total number of dyads in the network is (16 choose 2), or 120.

What does the network look like at different time points? The function `network.collapse` allows us to pull out the network at an instantaneous time point (with the argument `at`), while the function `network.extract` can pull out a given spell (with the arguments `onset` and `terminus`).

```
net500 <- network.collapse(stergm.sim.1,at=500)
net500
```

Network attributes:

```
vertices = 16
directed = FALSE
hyper = FALSE
loops = FALSE
multiple = FALSE
bipartite = FALSE
total edges= 15
  missing edges= 0
  non-missing edges= 15
```

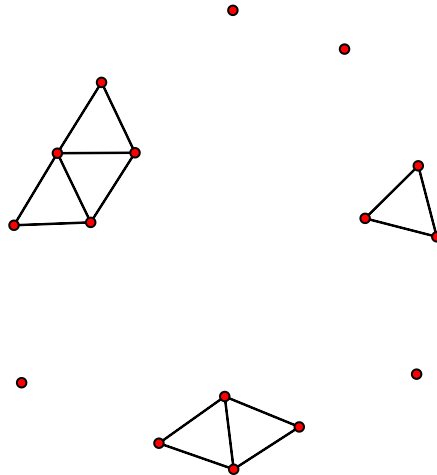
Vertex attribute names:

```
priorates totalties vertex.names wealth
```

No edge attributes

And we can look at the network structure:

```
plot(net500)
```



How well do the cross-sectional networks within our simulated dynamic network fit the probability distribution implied by our model? We can check by considering the summary statistics for our observed network, and those for our cross-sectional networks. This is easy, since by default, the `simulate` command for a `stergm` object not only simulates a dynamic network, but calculates the statistics from the model for each time point. These are stored in `attribute(simulated.networkDynamic)$stats`.

```
summary(flobusiness~edges+gwap(0,fixed=T))
```

```
edges gwap.fixed.0
  15          12
```

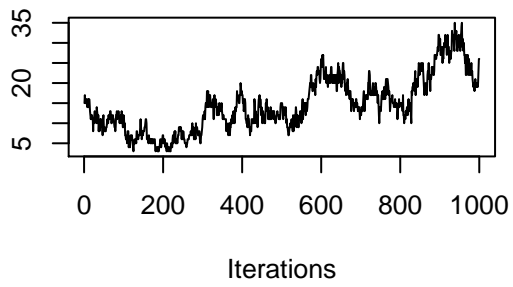
```
colMeans(attributes(stergm.sim.1)$stats)
```

```
edges gwap.fixed.0
14.675    11.752
```

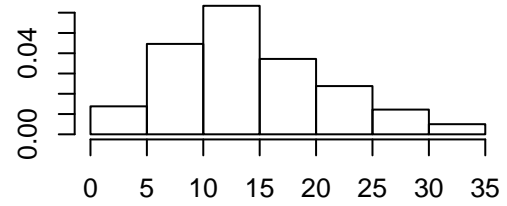
And we can also easily look at a time series and histogram for each statistic:

```
plot(attributes(stergm.sim.1)$stats)
```

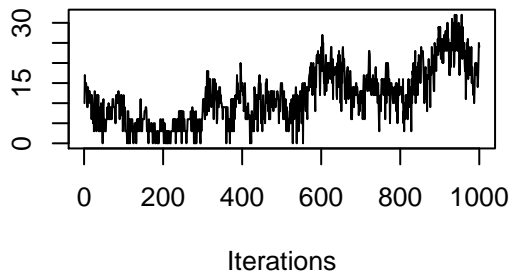
**Trace of edges**



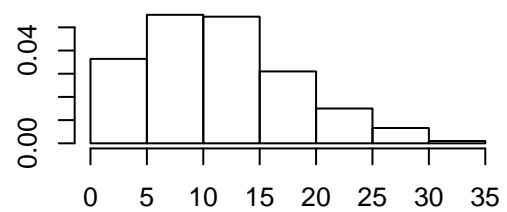
**Density of edges**



**Trace of gwesp.fixed.0**

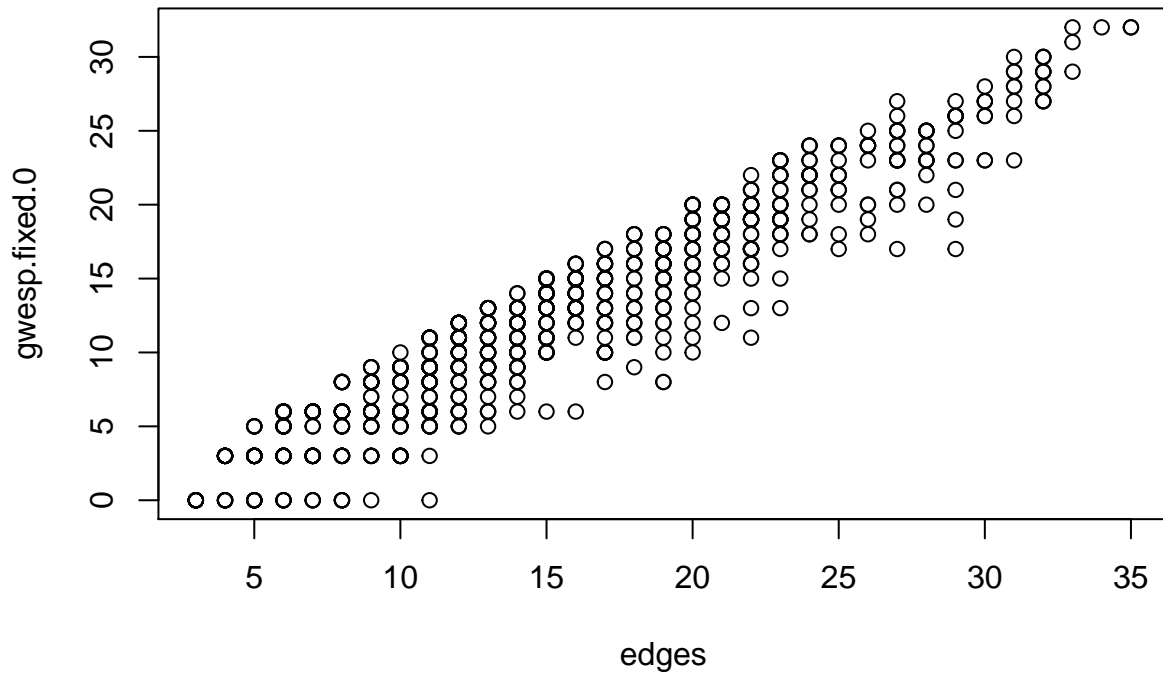


**Density of gwesp.fixed.0**



Or even at a scatter plot of the two network statistics for each simulated network cross-section, to see how these co-vary for this model:

```
plot(as.matrix(attributes(stergm.sim.1)$stats))
```



We should also check to make sure that our mean duration is what we expect (10 time steps). This requires knowing an additional function: `as.data.frame`, which, when applied to an object of class `networkDynamic`, generates a timed edgelist. Although right-censoring is present for some edges in our simulation, with a mean duration of 10 time steps and a simulation 1000 time steps long, its effect on our observed mean duration should be small.

```
stergm.sim.1.df <- as.data.frame(stergm.sim.1)
names(stergm.sim.1.df)
```

```
[1] "onset"           "terminus"       "tail"
[4] "head"           "onset.censored" "terminus.censored"
[7] "duration"       "edge.id"
```

```
stergm.sim.1.df[1,]
```

```
  onset terminus tail head onset.censored terminus.censored duration
1     0         3   3   5             TRUE                FALSE      3
edge.id
1     1
```

```
mean(stergm.sim.1.df$duration)
```

```
[1] 9.918974
```

The information on when an edge is active and when it is inactive is stored within our `network` object as the edge attribute `active`. Vertices, too, are capable of becoming active and inactive within `networkDynamic`, and this information is stored as a vertex attribute. Most of the time, users should access this information indirectly, through functions like `network.extract` or `as.data.frame`. Additional functions to query or set activity include `is.active`, `activate.vertex`, `deactivate.vertex`, `activate.edge`, and `deactivate.edge`, all documented in `help(package="networkDynamic")`.

Note that `networkDynamic` stores spells in the form `[onset,terminus)`, meaning that the spell is inclusive of the onset and exclusive of the terminus. So a spell of 3,7 means the edge begins at time point 3 and ends just before time point 7. `networkDynamic` can handle continuous-time spell information. However, since STERGMs are discrete-time with integer steps, what this means for STERGM is that the edge is inactive up through time step 2; active at time steps 3, 4, 5, and 6; and inactive again at time step 7 and on. Its duration is thus 4 time steps.

## 9. Visualizing dynamic networks using `ndtv`

The Network Dynamic Temporal Visualization (`ndtv`) package provides tools for visualizing changes in network structure and attributes over time. `ndtv` has its own tutorial at Sunbelt, with materials at <https://statnet.csde.washington.edu/trac/wiki/Sunbelt2016#WorkshopMaterials>

A quick taste of its functionality is here:

```
install.packages('ndtv')
library(ndtv)
stergm.sim.1a <- simulate.stergm(stergm.fit.1, nsim=1,
  time.slices = 100)
slice.par=list(start = 0, end = 25, interval=1, aggregate.dur=1, rule="any")
compute.animation(stergm.sim.1a, slice.par = slice.par)
render.par=list(tween.frames=5,show.time=T,
  show.stats="~edges+gwesp(0,fixed=T)")
wealthsize <- log(get.vertex.attribute(flobusiness, "wealth")) * 2/3
render.animation(stergm.sim.1a,render.par=render.par,
  edge.col="darkgray",displaylabels=T,
  label.cex=.8,label.col="blue",
  vertex.cex=wealthsize)
x11()
ani.replay()
```

## 10. Independence within and across time steps

STERGMs assume that the formation and dissolution processes are independent of each other *within the same time step*.

This does not necessarily mean that they will be independent across time. In fact, for any dyadic dependent model, they will not. To see this point, think of a romantic relationship example with:

```
formation = ~edges+degree(2:10)
dissolution = ~edges
```

with increasingly negative parameters on the degree terms.

What this means is that there is some underlying tendency for relational formation to occur, which is considerably reduced with each pre-existing tie that the two actors involved are already in. In other

words, there is a strong prohibition against being in multiple simultaneous romantic relationships. However, dissolution is fully independent—all existing relationships have the same underlying dissolution probability at every time step. (The latter assumption is probably unrealistic; in practice, if someone just acquired a second partner, their first is likely to be at increased risk of dissolving their relation. We ignore this now for simplicity).

Imagine that Chris and Pat are in a relationship at time  $t$ . During the time period between  $t$  and  $t+1$ , whether they break up does not depend on when either of them acquires a new partner, and vice versa. Let us assume that they do *not* break up during this time. Now, during the time period between  $t+1$  and  $t+2$ , whether or not they break up is dependent on the state of the network at time  $t+1$ , and that depends on whether either of them they acquired new partners between  $t$  and  $t+1$ .

The simple implication of this is that in this framework, formation and dissolution can be dependent, but that dependence occurs in subsequent time steps, not simultaneously.

Note that a time step here is arbitrary, and left to the user to define. One reason to select a smaller time interval is that it makes this assumption more justifiable. With a time step of 1 month, then if I start a new relationship today, the earliest I can break up with my first partner as a direct result of that new partnership is in one month. If my time step is a day, then it is in 1 day; the latter is likely much more reasonable. The tradeoff is that a shorter time interval means longer computation time for both model estimation and simulation, as will be seen below. You will see throughout this talk that there are multiple positives and negatives to having a short time step and having a long time step.

At the limit, this can in practice approximate a continuous-time model—the only issue is computational limitations.

## 11. Example 3: Approximation with long durations

For the type of model we saw in Example 2 (with a known dissolution model that contains a subset of terms from the formation model), it can be shown that a good set of starting values for the estimation of the formation model are as follows:

- (1) fit the terms in the formation model as a static ERGM on the cross-sectional network; and
- (2) subtract the values of the dissolution parameters from the corresponding values in the cross-sectional model. The result is a vector of parameter values that form a reasonable place to start the MCMC chain for the estimation of the formation model.

This is in fact exactly what the `stergm` estimation code does by default for this type of model.

When mean relational duration is very long, this approximation is so good that it may not be necessary to run a STERGM estimation at all. Especially if your purpose is mainly for simulation, the approximation may be all you need. This is a very useful finding, since models with long mean duration are precisely the ones that are the slowest and most difficult to fit using EGMME. That’s because, with long durations, very few ties will change between one time step and another, giving the fitting algorithm very little information on which to perform the estimation.

One way to frame this is to think of the classic approximation from epidemiology. Under some circumstances:

$$prevalence \approx incidence \times duration$$

which can be re-written as

$$incidence \approx prevalence \div duration$$

The same relationship that holds for levels of infection also holds for levels of relationships under some conditions (in this case, long relationships). The dissolution/persistence model gives us log-odds that map onto

duration. Fitting the cross-sectional ergm gives us log-odds for prevalence. We want to estimate incidence, so we subtract the two, rather than divide, since we are working on a log scale.

Of course, in order to be able to take advantage of this method, it is necessary for the terms in your dissolution model to be a subset of the terms in your formation model.

To illustrate, let us reconsider Example 2, with a mean relational duration of 100 time steps.

```
theta.diss.100 <- log(99)
```

First, we treat the formation process as if it were a stand-alone cross-sectional model, and estimate it using a standard cross-sectional ERGM. We did, in fact, fit this cross-sectional model earlier:

```
summary(fit1)
```

```
=====
Summary of model fit
=====

Formula:   flobusiness ~ edges + gwesp(0, fixed = T)

Iterations: 2 out of 20

Monte Carlo MLE Results:
              Estimate Std. Error MCMC % p-value
edges          -3.3626    0.6267      0 < 1e-04 ***
gwesp.fixed.0   1.5668    0.5927      0 0.00933 **
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

      Null Deviance: 166.36 on 120 degrees of freedom
Residual Deviance:  78.34 on 118 degrees of freedom

AIC: 82.34   BIC: 87.92   (Smaller is better.)
```

```
theta.form <- fit1$coef
theta.form
```

```
      edges gwesp.fixed.0
-3.362617    1.566777
```

Then, we subtract the values of the dissolution  $\theta$  from each of the corresponding values in the formation model. In this example, the dissolution model contains only an edges term, so this coefficient should be subtracted from the starting value for the edges term in the formation model.

```
theta.form[1] <- theta.form[1] - theta.diss.100
```

How well does this approximation do in capturing our desired dynamic network properties? First, we can simulate from it:



```
stergm.sim.2 <- simulate(flobusiness, formation=~edges+gwestp(0,fixed=T),
  dissolution=~edges, monitor="all",
  coef.form=theta.form, coef.diss=theta.diss.100,
  time.slices=50000)
```

Then check the results in terms of cross-sectional network structure and mean relational duration.

```
summary(flobusiness~edges+gwestp(0,fixed=T))
```

```
edges gwesp.fixed.0
 15      12
```

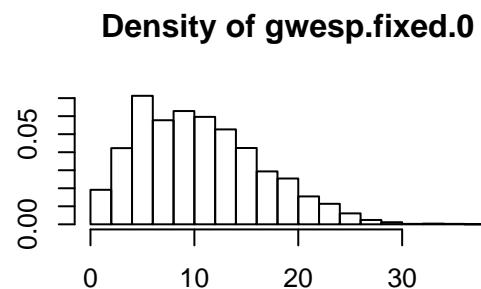
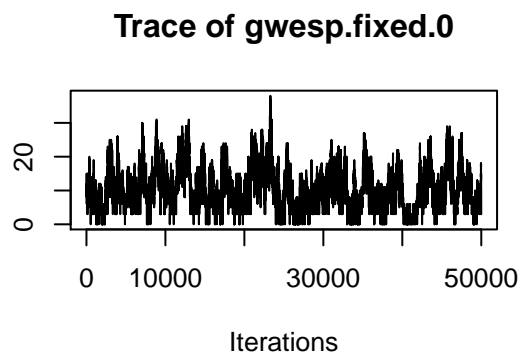
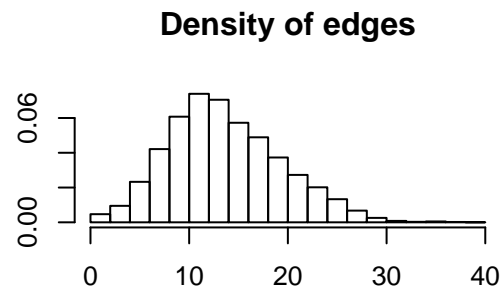
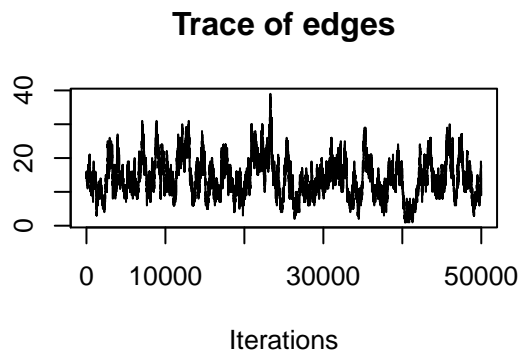
```
colMeans(attributes(stergm.sim.2)$stats)
```

```
edges gwesp.fixed.0
14.10646      11.03010
```

```
stergm.sim.dm.2 <- as.data.frame(stergm.sim.2)
mean(stergm.sim.dm.2$duration)
```

```
[1] 98.99481
```

```
plot(attributes(stergm.sim.2)$stats)
```



## 12. Example 4: Simulation driven by egocentric data

In many cases, people's primary interest in using dynamic networks is to simulate some diffusion process on one or more networks with similar features. Increasingly, our knowledge about those features come in the form of egocentrically sampled data, not from the traditional network census in a bounded population. Both `ergm` and `stergm` have methods for handling these situations.

For example, imagine that you want to model HIV transmission among a population of gay men in steady partnerships. 50% of the men are White and 50% are Black. You collect egocentric partnership data from a vaguely random sample of these men. Your data say:

- There are no significant differences in the distribution of momentary degree (the number of ongoing partnerships at one point in time) reported by White vs. Black men. The mean is 0.90, and the overall distribution is:
- 36% degree 0
- 46% degree 1
- 18% degree 2+
- 83.3% (i.e. 5/6) of relationships are racially homogeneous

We also have data (from these same men, or elsewhere) that tell us that the mean duration for a racially homogenous relationship is 10 months, while for a racially mixed one it is 20 months. Perhaps this is because the social pressure against cross-race ties makes it such that those who are willing to enter them are a select group more committed to their relationships.

Before we model the disease transmission, we need a dynamic network that possesses each of these features to simulate it on.

Our first step is to create a 500-node undirected network, and assign the first 250 nodes to race 0 and the second to race 1. The choice of 500 nodes is arbitrary.

```
msm.net <- network.initialize(500, directed=F)
msm.net %v% 'race' <- c(rep(0,250),rep(1,250))
msm.net
```

Network attributes:

```
vertices = 500
directed = FALSE
hyper = FALSE
loops = FALSE
multiple = FALSE
bipartite = FALSE
total edges= 0
  missing edges= 0
  non-missing edges= 0
```

Vertex attribute names:

```
race vertex.names
```

No edge attributes

ERGM and STERGM have functionality that allow us to simply state what the target statistics are that we want to match; we do not actually need to generate a network that has them. One way that we can specify the formation formula and target statistics is:

```
msm.form.formula <- ~edges+nodematch('race')+degree(0)+ concurrent
msm.target.stats <- c(225,187,180,90)
```

How did we get those values? Why don't we specify `degree(1)` as well?

Now let us turn to dissolution. We are back to the case where we can solve these explicitly, although this is complicated slightly by the fact that our dissolution probabilities differ by the race composition of the members. One dissolution formula for representing this is:

```
msm.diss.formula <- ~offset(edges)+offset(nodematch("race"))
```

These two model statistics means that there will be two model coefficients. Let us call them  $\theta_1$  and  $\theta_2$  for the edges and nodematch terms, respectively. Let us also refer to the change statistics for actor pair  $i, j$  for each of these as  $\delta(g_1(y))_{ij}$  and  $\delta(g_2(y))_{ij}$ , respectively.

Thus the log-odds expression for dissolution that we saw earlier would here be expressed as:

$$\ln \frac{P(Y_{ij,t+1}=1|Y_{ij,t}=1)}{P(Y_{ij,t+1}=0|Y_{ij,t}=1)} = \theta_1 \delta(g_1(y))_{ij} + \theta_2 \delta(g_2(y))_{ij}$$

which, in words, means that the conditional log-odds of a tie persisting equals  $\theta_1$  times the number of ties it involves, plus  $\theta_2$  times the number of race-homophilous ties it involves.

Note, then, that  $\delta(g_1(y))_{ij}$  would equal 1 for all actor pairs, while  $\delta(g_2(y))_{ij}$  would equal 1 for race homophilous pairs and 0 for others.

That means that the log-odds of tie persistence will equal  $\theta_1$  for mixed-race couples and  $\theta_1 + \theta_2$  for race-homophilous couples.

This suggests that we should be able to calculate  $\theta_1$  directly, and subsequently calculate  $\theta_2$ .

Following the logic we saw in Example 2, we can see that:

$$\theta_1 = \ln(d_{mixed} - 1)$$

and therefore

$$\theta_1 = \ln(20 - 1) = \ln 19 = 2.944$$

Furthermore,

$$\theta_1 + \theta_2 = \ln(d_{homoph} - 1)$$

and therefore

$$\theta_2 = \ln(d_{homoph} - 1) - \theta_1 = \ln(10 - 1) - 2.944 = -0.747.$$

So, we have:

```
msm.theta.diss <- c(2.944, -0.747)
```

We add in one additional control parameter—`SA.init.gain`—giving it a small value (the default is 0.1). As the help page for `control.stergm` sagely advises, ‘‘If the process initially goes crazy beyond recovery, lower this value.’’ This slows down estimation, but also makes it more stable. From trial and error, we know that this model, fit to this relatively large network, does better with this smaller value.

Putting it all together (including the syntax to control the window for real-time monitoring) gives us:

```
X11()
msm.fit <- stergm(msm.net,
  formation= msm.form.formula,
  dissolution= msm.diss.formula,
```

```

targets="formation",
target.stats= msm.target.stats,
offset.coef.diss = msm.theta.diss,
estimate = "EGMME",
control=control.stergm(SA.plot.progress=TRUE,
  SA.init.gain=0.005)
)
dev.off()

```

Iteration 1 of at most 20:

=====  
 ===== Lots of output snipped. =====

=====  
 ===== Phase 3: Simulate from the fit and estimate standard errors. =====

Let's first check to make sure it fit well:

```
mcmc.diagnostics(msm.fit)
```

```

=====
EGMME diagnostics
=====

```

Sample statistics summary:

```

Iterations = 10:2509
Thinning interval = 1
Number of chains = 1
Sample size per chain = 2500

```

1. Empirical mean and standard deviation for each variable,  
 plus standard error of the mean:

	Mean	SD	Naive SE	Time-series SE
edges	-0.1616	13.90	0.2781	1.1902
nodematch.race	-0.5752	12.83	0.2565	1.0460
degree0	0.6964	12.90	0.2580	0.8158
concurrent	-0.4344	10.69	0.2137	0.8911

2. Quantiles for each variable:

	2.5%	25%	50%	75%	97.5%
edges	-26	-10	-1	9	28
nodematch.race	-24	-10	-1	8	26
degree0	-24	-8	1	9	25
concurrent	-19	-8	-1	7	22

Sample statistics cross-correlations:

	edges	nodematch.race	degree0	concurrent
edges	1.0000000	0.8913476	-0.8320464	0.8743057

nodematch.race	0.8913476	1.0000000	-0.7321362	0.7887268
degree0	-0.8320464	-0.7321362	1.0000000	-0.5695312
concurrent	0.8743057	0.7887268	-0.5695312	1.0000000

Sample statistics auto-correlation:

Chain 1

	edges	nodematch.race	degree0	concurrent
Lag 0	1.0000000	1.0000000	1.0000000	1.0000000
Lag 1	0.8964313	0.8924016	0.8180669	0.8911414
Lag 2	0.8000610	0.7905685	0.6639729	0.7952635
Lag 3	0.7099515	0.6975145	0.5426190	0.7058012
Lag 4	0.6276052	0.6140555	0.4336958	0.6271920
Lag 5	0.5561281	0.5400089	0.3497867	0.5608359

Sample statistics burn-in diagnostic (Geweke):

Chain 1

Fraction in 1st window = 0.1

Fraction in 2nd window = 0.5

edges	nodematch.race	degree0	concurrent
0.06337	0.30569	-0.42428	-0.12856

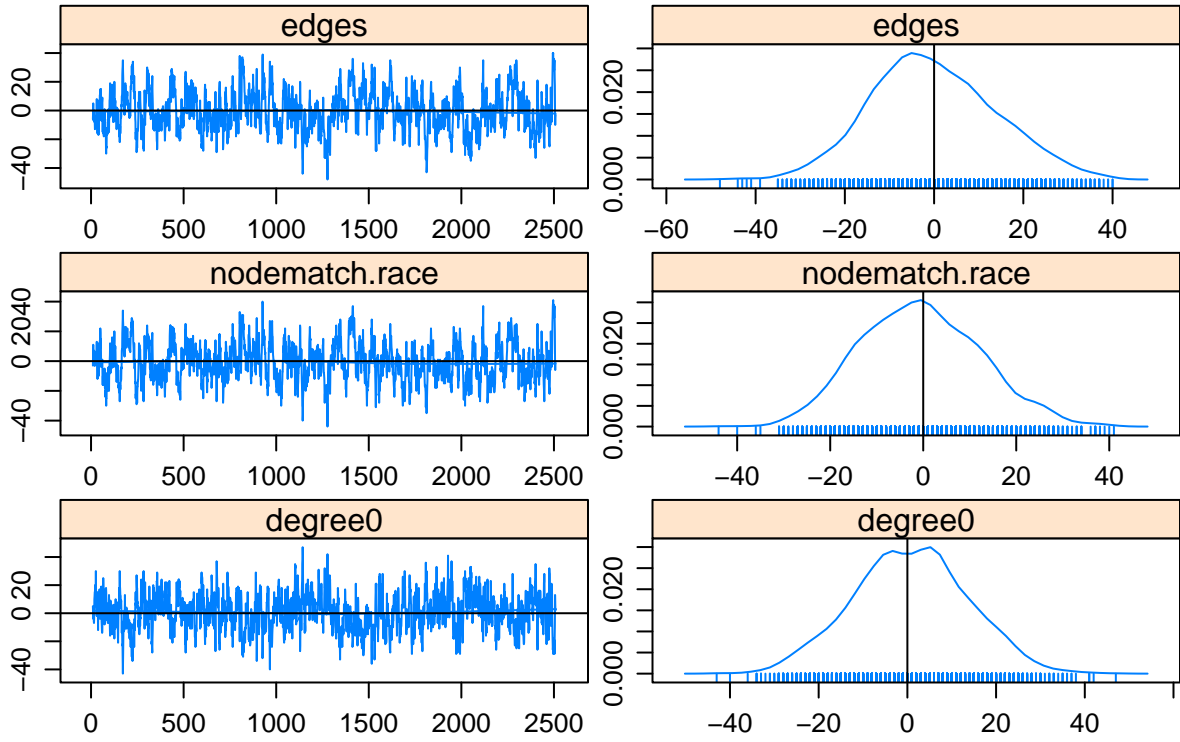
Individual P-values (lower = worse):

edges	nodematch.race	degree0	concurrent
0.9494685	0.7598419	0.6713580	0.8977086

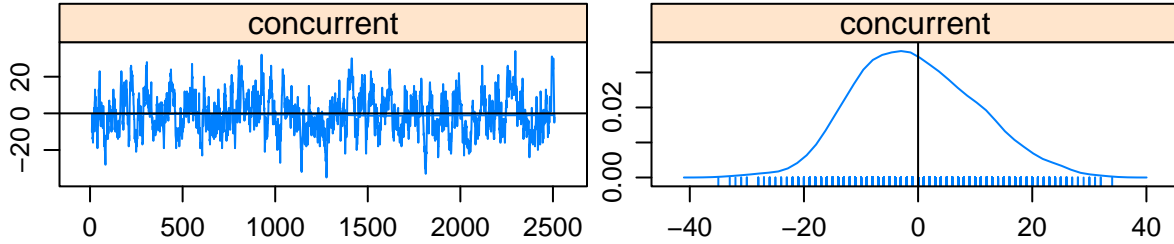
Joint P-value (lower = worse): 0.9065756 .

Loading required namespace: latticeExtra

# Sample statistics



## Sample statistics



MCMC diagnostics shown here are from the last round of simulation, prior to computation of final parameters

and see what the results tell us:

```
summary(msm.fit)
```

```
=====
Summary of formation model fit
=====
```

```
Formula: ~edges + nodematch("race") + degree(0) + concurrent
```

```
Iterations: NA
```

```
Equilibrium Generalized Method of Moments Results:
```

	Estimate	Std. Error	MCMC %	p-value
edges	-9.9328	0.4255	0	< 1e-04 ***
nodematch.race	2.2864	0.2338	0	< 1e-04 ***
degree0	-0.1879	0.1138	0	0.098790 .
concurrent	-0.8431	0.2400	0	0.000443 ***

```
---
```

```
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
=====
Summary of dissolution model fit
=====
```

```
Formula: ~offset(edges) + offset(nodematch("race"))
```

```
Iterations: NA
```

```
Equilibrium Generalized Method of Moments Results:
```

	Estimate	Std. Error	MCMC %	p-value
edges	2.944	0.000	0	<1e-04 ***
nodematch.race	-0.747	0.000	0	<1e-04 ***

```
---
```

```
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

The following terms are fixed by offset and are not estimated:  
edges nodematch.race

One should not over-interpret the standard errors and p-values in this case. Remember, the size of the population that we built to fit this model was arbitrary (although it could have been constructed to match the source data). Our main goal in fitting the model is to be able to simulate a dynamic network with both desired cross-sectional and durational features maintained stochastically over time.

Now, to do that, we simulate a dynamic network:

```
msm.sim <- simulate(msm.fit,time.slices=1000)
```

and compare the outputs to what we expect, in terms of cross-sectional structure:

```
colMeans(attributes(msm.sim)$stats)
```

edges	nodematch.race	degree0	concurrent
226.136	187.710	180.333	90.989

```
msm.target.stats
```

```
[1] 225 187 180 90
```

And relationship length:

```
race <- msm.net %v% 'race'
msm.sim.dm <- as.data.frame(msm.sim)
names(msm.sim.dm)
```

[1] "onset"	"terminus"	"tail"
[4] "head"	"onset.censored"	"terminus.censored"
[7] "duration"	"edge.id"	



```
mean(msm.sim.dm$duration[race[msm.sim.dm$tail] == race[msm.sim.dm$head]])
```

```
[1] 10.00037
```

```
mean(msm.sim.dm$duration[race[msm.sim.dm$tail] != race[msm.sim.dm$head]])
```

```
[1] 19.79619
```

Don't run this in real-time during the tutorial, as it is slow, but if you wished to visualize this dynamic network, the following code would do so:

```
slice.par=list(start = 0, end = 100, interval=1, aggregate.dur=1, rule="any")
compute.animation(msm.sim, slice.par = slice.par)

render.par=list(tween.frames=5,show.time=T)
render.animation(msm.sim,render.par=render.par,
  edge.col="darkgray", displaylabels=F, vertex.col="race")
x11()
ani.replay()
```

### 13. Additional functionality

Both the `stergm` functions and the `networkDynamic` package have additional functionality, which you can learn about and explore through the use of R's many help features.

If you begin to use them in depth, you will likely have further questions. If so, we encourage you to join the statnet users' group ([http://csde.washington.edu/statnet/statnet\\_users\\_group.shtml](http://csde.washington.edu/statnet/statnet_users_group.shtml)), where you can then post your questions (and possibly answer others). You may also encounter bugs; please use the same place to report them.

### References

- Carter T. Butts, Ayn Leslie-Cook, Pavel N. Krivitsky, and Skye Bender-deMoll. `networkDynamic`: Dynamic Extensions for Network Objects. The Statnet Project <http://www.statnet.org>, 2013. R package version 0.6. <http://CRAN.R-project.org/package=networkDynamic>
- Krivitsky, P.N., Handcock, M.S.(2014). A separable model for dynamic networks *JRSS Series B-Statistical Methodology*, 76 (1):29-46; 10.1111/rssb.12014 JAN 2014
- Pavel N. Krivitsky. Modeling of Dynamic Networks based on Egocentric Data with Durational Information. Pennsylvania State University Department of Statistics, 2012(2012-01). <http://stat.psu.edu/research/technical-reports/2012-technical-reports>
- Pavel N. Krivitsky and Mark S. Handcock. `tergm`: Fit, Simulate and Diagnose Models for Network Evolution based on Exponential-Family Random Graph Models. The Statnet Project <http://www.statnet.org>, 2013. R package version 3.1-0. <http://CRAN.R-project.org/package=tergm>